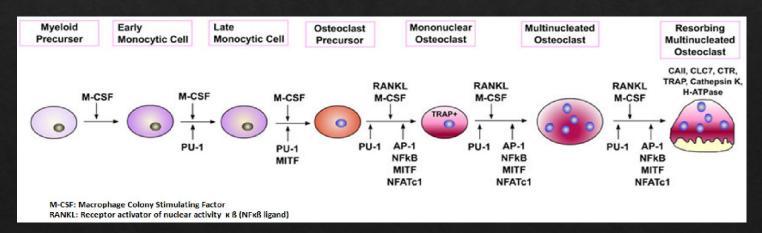


Sweet Surprises: Unraveling the Sugary Side of Notch and How Overexpressed Fringe Enzymes Shape Osteoclastogenesis

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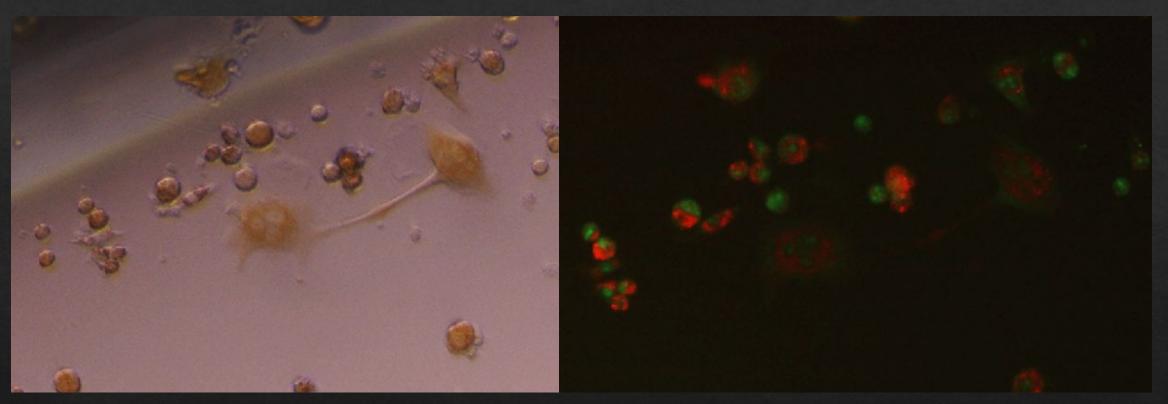
Background Information

- Osteoclast cells are large multinucleated phagocytic cells that play a crucial role in the resorption or breakdown of bone tissue in addition to their role in bone remodeling, growth, and repair.
- * **Osteoclastogenesis** is dependent on RANK/RANKL pathway signaling while other pathways, such as Notch signaling, influence other factors of the cell life such as size, activity, and lifespan.
- Fringe enzymes are a specific N-acetylglucosaminyltransferase that extends O- fucose monosaccharides which modulates Notch pathway activity. Within mammals there are 3 homologs: Lunatic, Manic and Radical Fringes.



Stages of osteoclast differentiation from lineage cells [Soltanoff, et al. 2009]

Osteoclast Are Multinucleated Cells



Brightfield Staining

Acridine Orange

Experiment Overview

Experiment 1

- Impact of time on osteoclast formation within RAW264.7 cell lines [cultures fixed at 3 or 4 days after exposure to RANKL]
- Control: EFGP fluorescing plasmids or M-Cherry fluorescing plasmids
- Experiment 2
 - Overexpression of Fringe glycosyltransferase within mouse BMM differentiation
 - I Plating methods: Direct seeding into 96 well plates or transferring from 60mm petri dishes to 96 wells
 - Direct seeding is less stressful to the delicate precursor cells but does not allow for accurate cell numbers
 - * 60mm transferred methodology tested by exposing virus for 24hrs or 6 hrs.

Methodology Overview

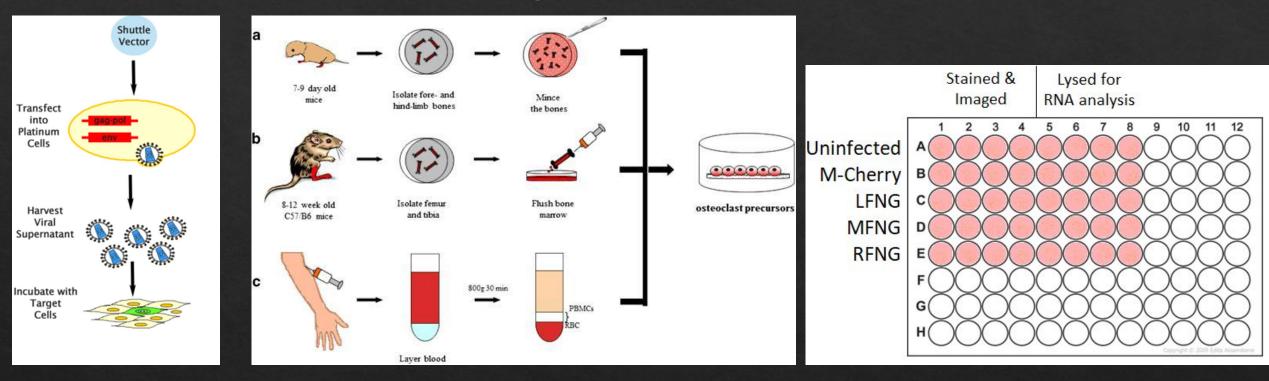
Creating Overexpression:

Specific plasmids when introduced to a macrophage can overexpress (OE) Fringe enzymes. Viral plasmids are created with PLAT-E packaging cells infected with cDNA and membrane transgenes. After 48hrs the media supernatant is collected and stored for later infection of macrophages.

♦ Cell Lines:

- RAW264.7 are immortal mouse cells established from a male mouse tumor. cultured the cell lines for several passages, infected with plasmids, selected with puromycin antibiotic, seeded into 96 well plates, then differentiated with RANKL. After cells are stained with Acridine Orange, fixed and then stained for TRAP (Tartrate Resistant Acid Phosphate) activity.
- Primary bone marrow macrophages are mouse cells collected from male mice femurs and tibias, isolated through culturing techniques, seeded in either 96 well plates (50,000 cells/well) or 60 mm petri dishes with M-CSF, transfected with viral plasmids, 60mm cultures seeded into 96 well plates with (10,000 cells/well), selected with puromycin antibiotic, and differentiated with RANKL.

Methodology Overview



Plasmids within the viral supernatant are crystallized within calcium phosphate microscopic crystals to shuttle plasmids within the macrophages to select for OE transgenes. [Cell Biolabs, 2008] Osteoclast precursor collection methods vary across species analyzed. In animal studies the precursors are collected from the bone marrow directly. [Marino, et al. 2014] Diagram of 96-well plate used to analyze effectiveness of osteoclastogenesis within primary BMM

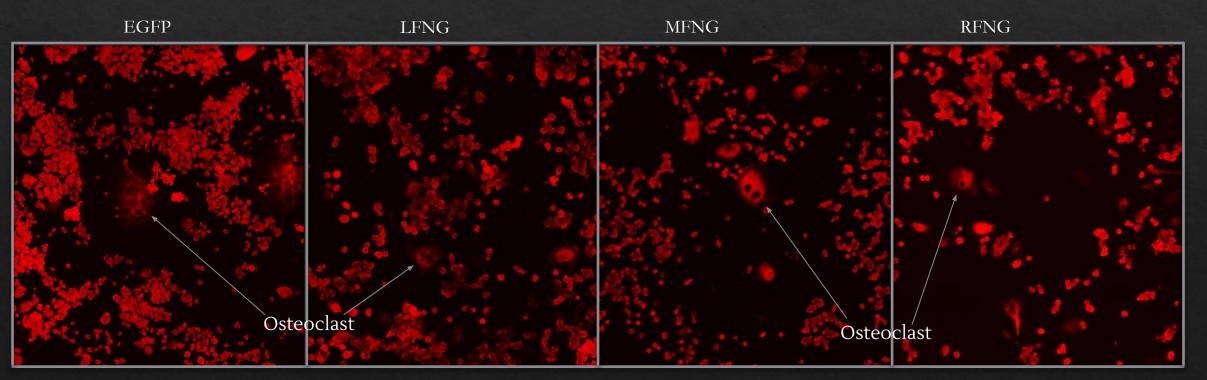
Results of Experiment 1

3 Days

- Osteoclastogenesis is reduced in all cultures
- Osteoclasts that did form are smaller with less nuclei

4 Days

- Osteoclasts increase in number within culture
- Larger osteoclasts with more nuclei, morphological differences within Fringe lines

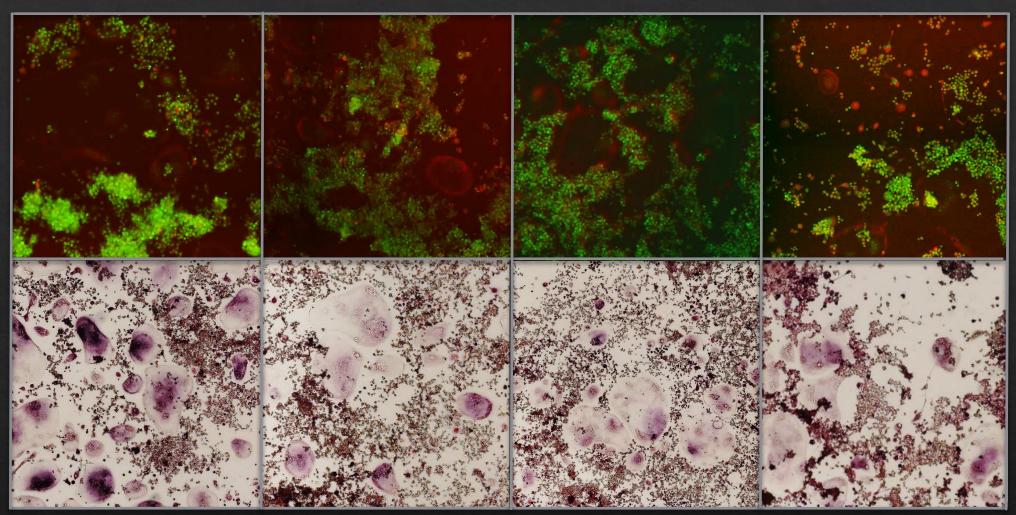


TRAP under Fluorescence (Texas Red) at 3 Days after RANKL exposure to induce differentiation



LFNG

MFNG



Control (EGFP)

LFNG: large active osteoclasts at time of fixing

MFNG: larger osteoclasts with reduced function at time of staining indicating early cellular death

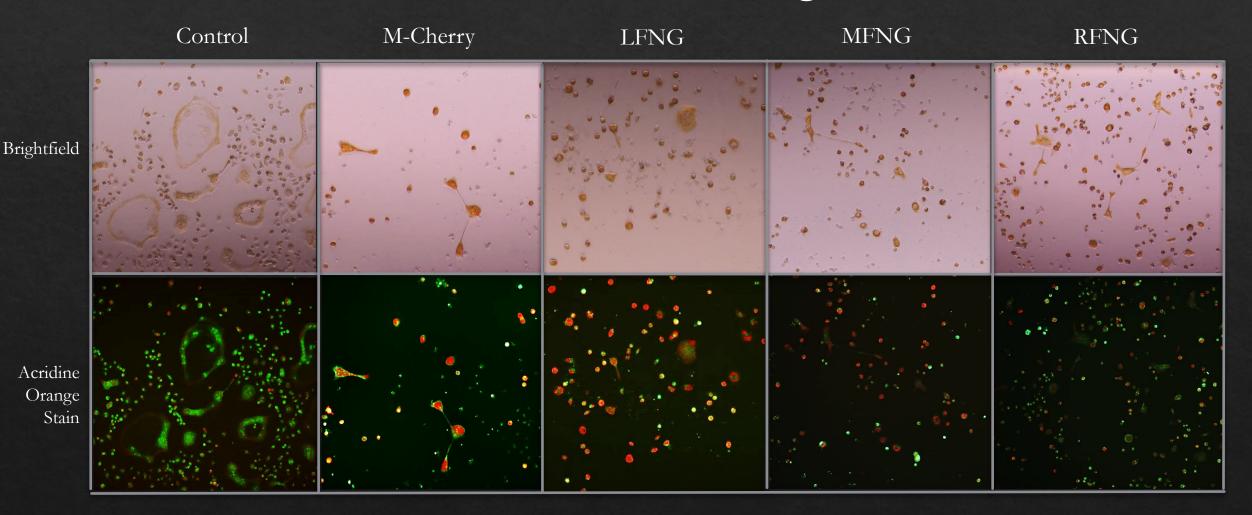
RFNG: fewer osteoclasts that presented smaller

AO

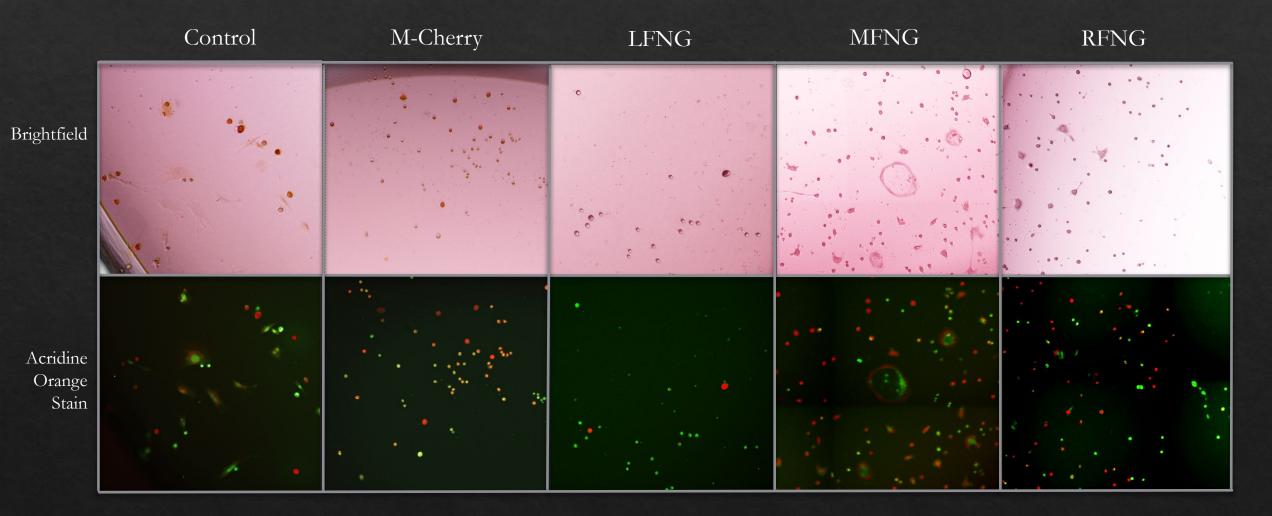
TRAP

Results of Experiment 2

96 well direct seeding



60mm Transfer to 96-Well



Discussion

Experiment 1

- Fringe homologs impact cell lifespan and morphology
- I Fluorescent plasmids as control interfere with staining.
- Experiment 2
 - Directly seeding BMM cells into 96 well plates and then transfecting showed the most notable impact to survival of transfection process with the densities tested
 - Exposing virus for 6 hours preserved the naïve state of the precursor macrophages, but alternative methods must be explored for higher survival rates during the transfer to 96 well plates
- What's Next?
 - □ Analysis of collected RNA from primary BMM to quantify Fringe OE
 - Test varying densities of direct seeding to 96 well plates
 - □ Knockout of Fringe homologs within RAW264.7 and primary BMM cell lines

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Selected References

- Soltanoff, C. *et al.* (2009). Signaling Networks that Control the Lineage Commitment and Differentiation of Bone Cells. Critical reviews in eukaryotic gene expression. 19. 1-46. 10.1615/CritRevEukarGeneExpr.v19.i1.10.
- Marino S, Logan JG, Mellis D, Capulli M. Generation and culture of osteoclasts. Bonekey Rep. 2014 Sep 10;3:570. doi: 10.1038/bonekey.2014.65
- LeBon, L. *et al.* (2014). Fringe proteins modulate Notch-ligand cis and trans interactions to specify signaling states.
 eLife, 3:e02950. https://doi.org/10.7554/eLife.02950
- Ashley, J. *et al.* (2015). Notch signaling promotes osteoclast maturation and resorptive activity. *J Cell Biochem*. 116(11):2598-609. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4565736/
- Morita, S. *et al.* (2000). Plat-E: an efficient and stable system for transient packaging of retroviruses. *Gene Ther*, 7, 1063–1066. https://doi.org/10.1038/sj.gt.3301206
- Maetzig, T., *et al.* (2011). Gammaretroviral vectors: biology, technology and application. *Viruses*, 3(6), 677–713. https://doi.org/10.3390/v3060677
- Kitamura, T. *et al.* (2003). Retrovirus-mediated gene transfer and expression cloning: powerful tools in functional genomics. *Experimental Hematology*, *31*, 1007-1014. https://doi.org/10.1016/j.exphem.2003.07.005.