



The background image shows a laboratory environment. In the foreground, there are several petri dishes containing bacterial cultures. One dish on the left has a streaked culture with handwritten labels 'J.I. 1243', 'ΔPlac', 'BL', and '7/17'. Another dish on the right shows a dense, dark purple bacterial growth with similar labels. Below the dishes is a blue microplate with wells, and an orange microplate is partially visible at the bottom. A white centrifuge tube rack is in the upper right, with a label that says 'Patented', 'Fold Flat to Recycle', and 'U.S. Patent #5,721,111'.

JANTHINOBACTERIUM LIVDUM

Monique's summer research project

Mentor : Dr. Ben Lungdren

KEY WORDS & EXPLANATION

RPON

A process in which scientist can learn how bacteria make things they need to survive and grow.

GRAM NEGATIVE

A tougher bacteria with a protective layer and prone to turn red/ pink. Used to tell what kind of bacteria it is

GENE TRANSMISSION

Genetic information passed to offspring

GENE EXPRESSION

How gene can control the way they are expressed or placed when pressured to adapt to environment.

ENVIRONMENTAL STRESS

a concept in which you put pressure

VIOLACEIN

A defense mechanism to fight off bacteri, fungi

AUTOClave

Used to sterilize, media, water, and clean tools

PLASMID

Tiny piece of DNA

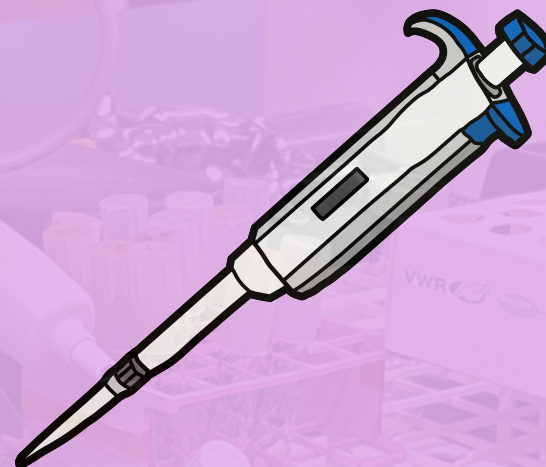
AGENDA



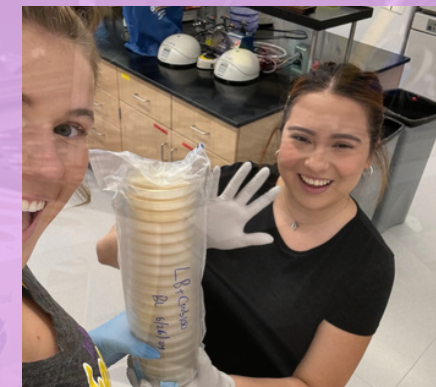
Abstract



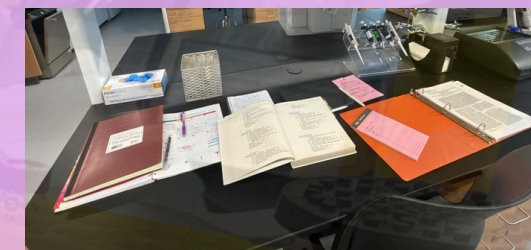
Background



methods/ limitations



**What's next &
acknowledgments**



References

QUESTIONS AT THE END !!

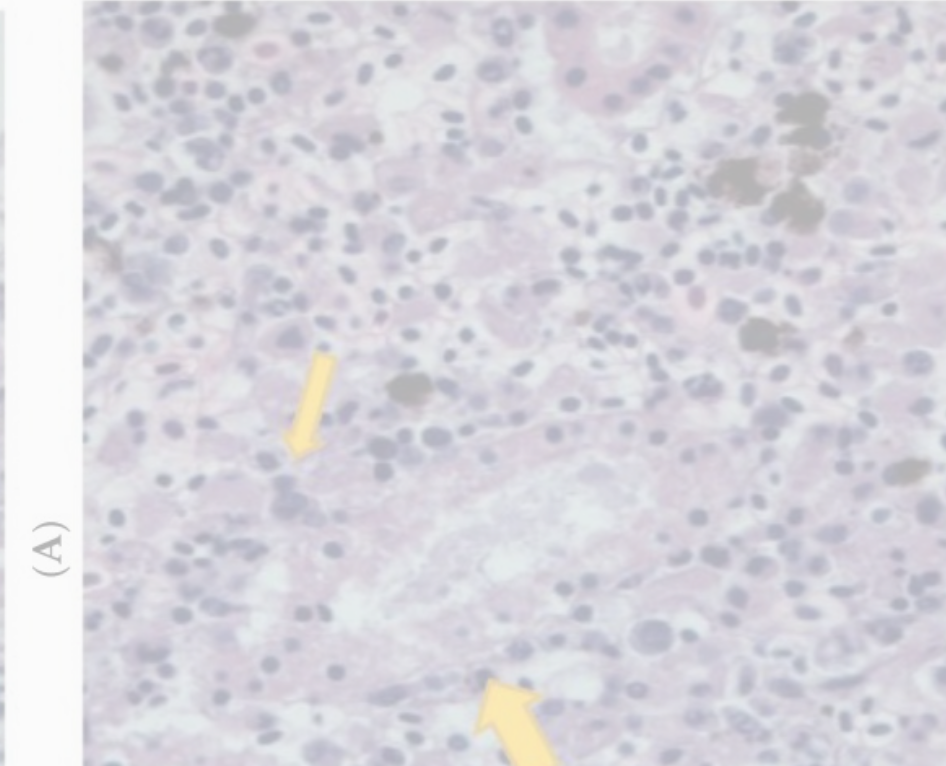
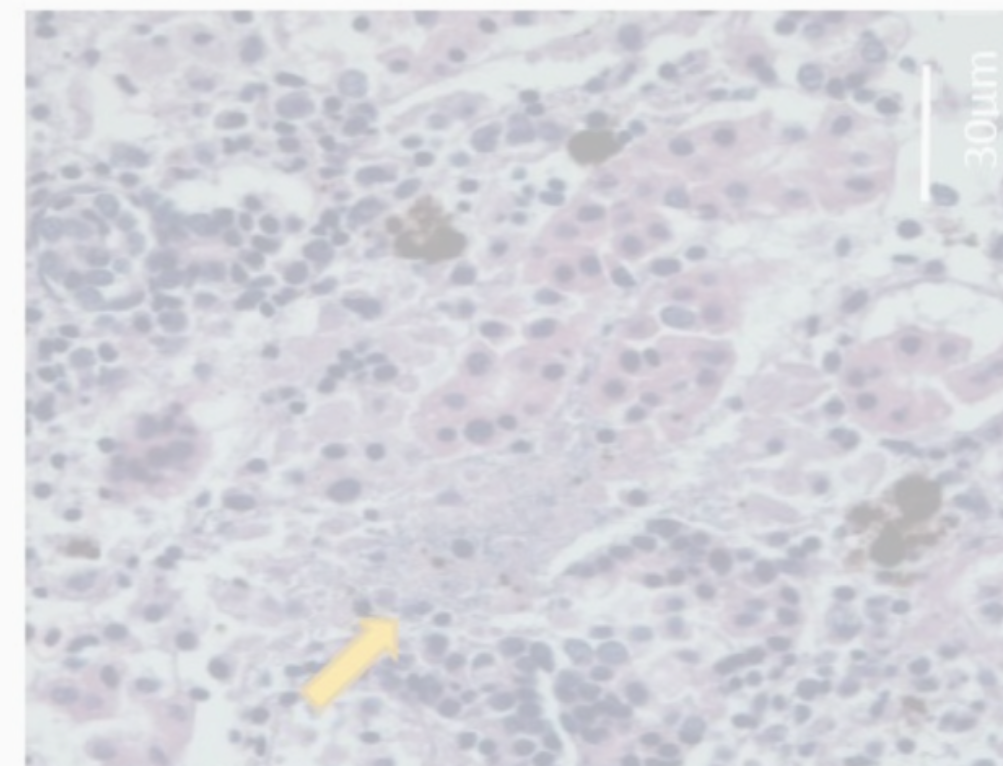
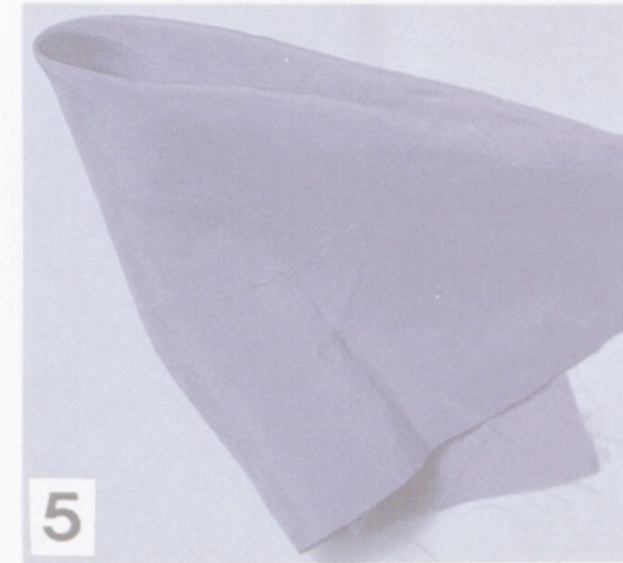
ABSTRACT / RESEARCH QUESTION

This research aims to understand and identify the role of RpoN within the adaptability of *Janthinobacterium* strain 12473 and the implications in violacein production. It's critical to analyze *J.lividum* to see which nitrogen source, *J.lividum*, can exponentially grow the best and emit violacein under experimental stress conditions. The methodology used in this research was quantitative and qualitative measures. I looked into its resistance by data recording and observations while referencing research articles that also share similar antibiotic testing methods, growth conditions, and observations of the samples throughout lac assays, culture growth of plates and other experiments. RpoN contributes to the understanding of environmental stress on *Janthinobacterium lividum* and can give insight how we can potentially use this to understand the limits of violacein. Furthermore, using this data we can address that *janthinobacterium* holds a key in how we can produce and implement violacein findings to help further research in medical applications with violacein.

BACKGROUND ON *J. LIVIDUM*

Janthinobacterium lividum, a gram negative soil-dwelling bacterium commonly found in lakes, soil, rivers, and arctic glaciers. *J. lividum* has become more prevalent in science research because of the interesting research studies it's involved in. One of the studies *J. lividum* has negatively affected trout industries in Korea, a popular food source in Asian countries. The research focused on the infection process the bacteria had on the rainbow trout. However, in another study *J. lividum* produces violacein that emits a purple color when fighting off fungi, and bacteria. Violacein is an important byproduct of this bacteria that aids in medical applications. An article published by American Society for Microbiology, states the strain 12473 of *J. lividum* helps in biosynthesis of the antibiotic daptomycin and anti-cancer compound epothilone.

In Addition, the violacein production of *J. lividum* is affected by environmental stress in global locations, it is found. Meaning whether it's located in The Americas, Korea, and Europe or Antarctica, environmental factors affect the strain by salinity levels, soil components, temperature the strain is found in. Therefore, Knowing this it's important to study and continually test this adaptive bacteria's resistance, metabolism and growth conditions when fighting off bacteria and fungi. Therefore, by using RpoN we can understand how *janthinobacterium lividum* regulates its gene expression enabling the transcription of certain genes, particularly those involved in nitrogen metabolism and induction of violacein.



METHODS USED:

Lac assays / plate streaking
abs buffer making / excel
defending / autoclave
filtration/electroporation and electrophoresis
media culture
biofilm prep

The quantitative data I analyzed a table of an absorbance spectrum of my samples at 420 & 600 comparing KNO₃ and NH₄Cl interactions with a *J.* plasmid.

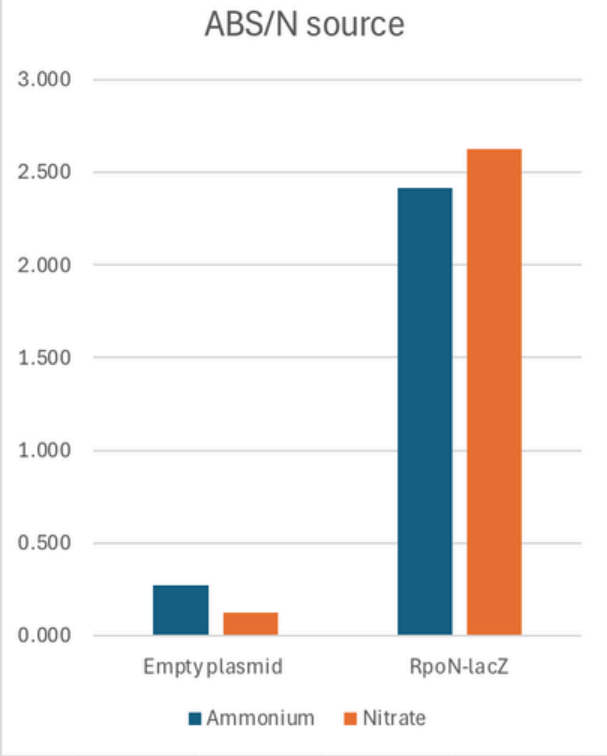
Quantitative data in my research also looks at positive or negative outcomes on growth of bacteria or emission of the color purple which means that violacein is working and also keeping note of the amount of grams I used while creating my own stocks and media.

Some of the qualitative data while studying *J. lividum*, noting observations of color and taking pictures and videos throughout my research experiment to note how I observed results and performed techniques.

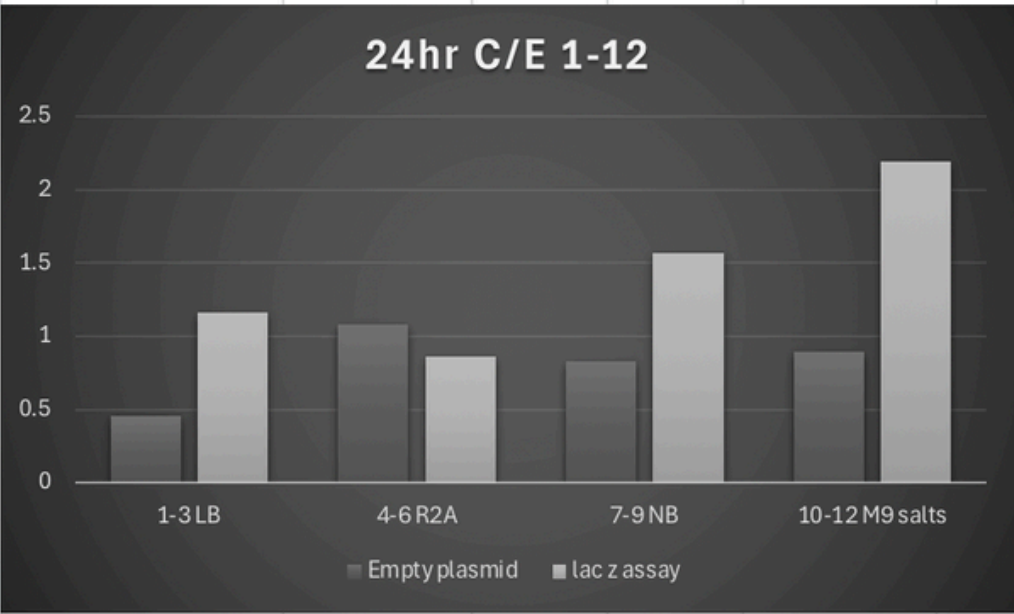


Data tables

	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P
1	Sample #	Abs 600	Actual Abs 6	Srt (min)	End (min)	RXN time (Abs 420	Miller unit	Miller mean ur	STRD DIV						
2	M1	0.261	1.57	1.5	1337.5	1336	0.622	2.973	2.415	0.53776709						
3	M2	0.130	0.780	1.5	1337.5	1336	0.198	1.900	2.415	0.53776709						
4	M3	0.162	0.972	1.5	1337.5	1336	0.308	2.372	2.415	0.53776709						
5	M4	0.230	1.38	0.5	1336.5	1336	0.657	3.564	2.622	0.85393108						
6	M5	0.236	1.42	0.5	1336.5	1336	0.359	1.898	2.622	0.85393108						
7	M6	0.193	1.16	0.5	1336.5	1336	0.372	2.405	2.622	0.85393108						
8	R1	0.210	1.26	13.5	1340.5	1327	0.037	0.221	0.269	0.05930713						
9	R2	0.116	0.696	13.5	1340.5	1327	0.031	0.336	0.269	0.05930713						
10	R3	0.155	0.930	13.5	1340.5	1327	0.031	0.251	0.269	0.05930713						
11	R4	0.270	1.62	13.0	1338.5	1325.5	0.016	0.075	0.123	0.05612797						
12	R5	0.143	0.858	13.0	1338	1325	0.021	0.185	0.123	0.05612797						
13	R6	0.249	1.49	13.0	1338	1325	0.022	0.111	0.123	0.05612797						
14																
15																
16	KEY	KEY			Empty plasm	RpoN-lacZ		STDEV E-pla	STDEV lac							
17	KNO3	PBR1 1007		Ammonium	0.269	2.415		0.0593071	0.53776709							
18	NH4Cl	PBBR MSC-5		Nitrate	0.123	2.622		0.056128	0.85393108							
19																
20																
21																

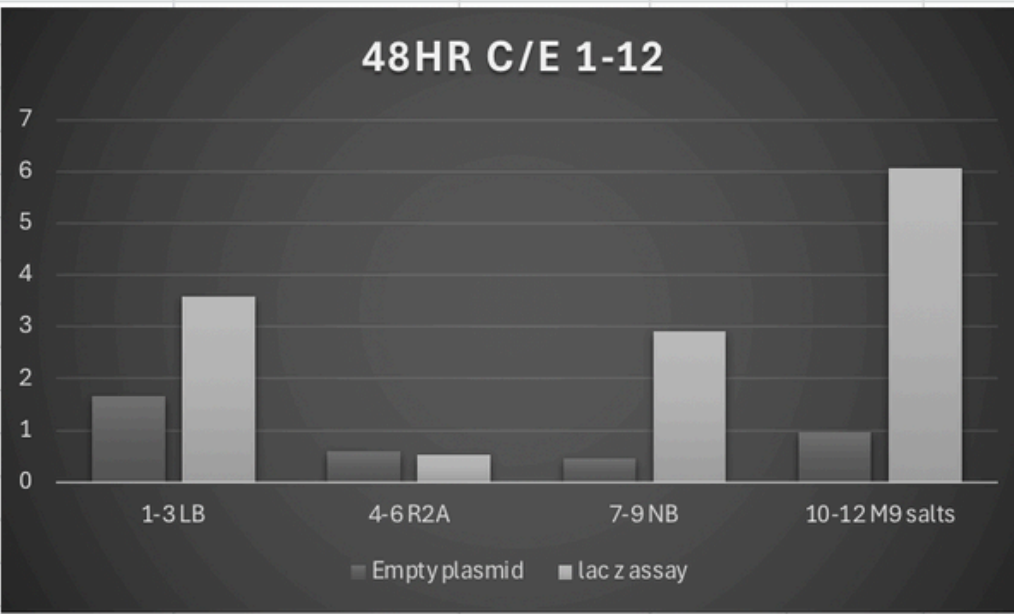


52						
53						
54						
55		24 hr) C/E	Empty plasmid	lac z assay	STDEV E-plasmid	STDEV lacZ
56		1-3 LB	0.458046698	1.15820572	0.179892526	0.510677433
57		4-6 R2A	1.077480779	0.860362553	0.369587095	0.37580940
58		7-9 NB	0.828254858	1.566486526	0.175675079	0.522243362
59		10-12 M9 salts	0.890422181	2.192038532	0.223444980	1.437624475
60						
61						
62						
63						
64						
65						
66						
67						
68						
69						
70						
71		48 hr) C/E	Empty plasmid	lac z assay	STDEV E-plasmid	STDEV lacZ
72		1-3 LB	1.663876351	3.577226618	0.373650514	1.372109119
73		4-6 R2A	0.591348803	0.519973984	0.198019455	0.155906124
74		7-9 NB	0.463967324	2.913953664	0.022533732	0.847926393
75		10-12 M9 salts	0.952623189	6.058897015	0.197478494	3.128089222
76						
77						
78						
79						



	A	B	C	D	E	F	G	H	I	J
1	Sample	Abs 600	Act abs 600	RXN Srt (min)	RXN end (min)	RXN time mean	Abs 420	Miller units	Miller mean units	STRD DIV
2	24) c1	0.071	0.852	1.0	1394	1393.0	0.079	0.665635353	0.458046698	0.179892526
3	24) c2	0.068	0.816	1.0	1394	1393.0	0.041	0.360697043	0.458046698	0.179892526
4	24) c3	0.086	1.032	1.0	1394	1393.0	0.050	0.347807699	0.458046698	0.179892526
5	24) c4	0.017	0.204	1.0	1394	1393.0	0.031	1.090888617	1.077480779	0.369587095
6	24) c5	0.029	0.348	1.0	1394	1393.0	0.034	0.701372214	1.077480779	0.369587095
7	24) c6	0.027	0.324	1.0	1394	1393.0	0.065	1.440181507	1.077480779	0.369587095
8	24) c7	0.039	0.468	1.0	1394	1393.0	0.047	0.720942932	0.828254858	0.175675079
9	24) c8	0.040	0.480	1.0	1394	1393.0	0.049	0.732830821	0.828254858	0.175675079
10	24) c9	0.047	0.564	1.0	1394	1393.0	0.081	1.03099082	0.828254858	0.175675079
11	24) c10	0.049	0.588	1.0	1394	1393.0	0.086	1.049953363	0.890422181	0.223444980
12	24) c11	0.065	0.780	1.0	1394	1393.0	0.069	0.635043349	0.890422181	0.223444980
13	24) c12	0.037	0.444	1.0	1394	1393.0	0.061	0.98626963	0.890422181	0.223444980
14	24) E1	0.084	1.008	3.0	1398	1395.0	0.122	0.867611083	1.15820572	0.510677433
15	24) E2	0.081	0.972	3.0	1398	1395.0	0.237	1.74786495	1.15820572	0.510677433
16	24) E3	0.089	1.068	3.0	1398	1395.0	0.128	0.859141127	1.15820572	0.510677433
17	24) E4	0.030	0.360	3.0	1398	1395.0	0.065	1.294305058	0.860362553	0.375809401
18	24) E5	0.075	0.900	3.0	1398	1395.0	0.081	0.64516129	0.860362553	0.375809401
19	24) E6	0.054	0.648	3.0	1398	1395.0	0.058	0.641621311	0.860362553	0.375809401
20	24) E7	0.076	0.912	3.0	1398	1395.0	0.123	0.966798717	1.566486526	0.522243362
21	24) E8	0.037	0.444	3.0	1398	1395.0	0.119	1.921276115	1.566486526	0.522243362
22	24) E9	0.031	0.372	3.0	1398	1395.0	0.094	1.811384746	1.566486526	0.522243362
23	24) E10	0.054	0.648	3.0	1398	1395.0	0.102	1.128368512	2.192038532	1.437624475
24	24) E11	0.027	0.324	3.0	1398	1395.0	0.173	3.827602991	2.192038532	1.437624475
25	24) E12	0.066	0.792	3.0	1398	1395.0	0.179	1.620144093	2.192038532	1.437624475
26	48) C1	0.087	1.044	93.5	1401	1307.5	0.266	1.948675121	1.663876351	0.373650514
27	48) C2	0.087	1.044	93.5	1401	1307.5	0.246	1.802158194	1.663876351	0.373650514
28	48) C3	0.094	1.128	93.5	1401	1307.5	0.183	1.240795737	1.663876351	0.373650514
29	48) C4	0.061	0.732	93.5	1401	1307.5	0.047	0.491071895	0.591348803	0.198019455
30	48) C5	0.066	0.792	93.5	1401	1307.5	0.048	0.463526276	0.591348803	0.198019455
31	48) C6	0.035	0.420	93.5	1401	1307.5	0.045	0.819448238	0.591348803	0.198019455
32	48) C7	0.067	0.804	93.5	1401	1307.5	0.047	0.447095307	0.463967324	0.022533732
33	48) C8	0.070	0.840	93.5	1401	1307.5	0.050	0.455249021	0.463967324	0.022533732
34	48) C9	0.069	0.828	93.5	1401	1307.5	0.053	0.489557643	0.463967324	0.022533732
35	48) C10	0.082	0.984	93.5	1401	1307.5	0.151	1.173654184	0.952623189	0.197478494
36	48) C11	0.078	0.936	93.5	1401	1307.5	0.109	0.890653854	0.952623189	0.197478494
37	48) C12	0.102	1.224	93.5	1401	1307.5	0.127	0.793561529	0.952623189	0.197478494
38	48) E1	0.093	1.116	97.0	1406	1309.0	0.753	5.15455449	3.577226618	1.372109119
39	48) E2	0.108	1.296	97.0	1406	1309.0	0.495	2.9178338	3.577226618	1.372109119
40	48) E3	0.079	0.948	97.0	1406	1309.0	0.330	2.659291565	3.577226618	1.372109119
41	48) E4	0.067	0.804	97.0	1406	1309.0	0.047	0.446582975	0.519973984	0.155906124
42	48) E5	0.051	0.612	97.0	1406	1309.0	0.056	0.699031841	0.519973984	0.155906124
43	48) E6	0.063	0.756	97.0	1406	1309.0	0.041	0.414307137	0.519973984	0.155906124
44	48) E7	0.057	0.684	97.0	1406	1309.0	0.299	3.339453804	2.913953664	0.847926393
45	48) E8	0.061	0.732	97.0	1406	1309.0	0.332	3.464873282	2.913953664	0.847926393
46	48) E9	0.069	0.828	97.0	1406	1309.0	0.210	1.937533907	2.913953664	0.847926393
47	48) E10	0.072	0.864	97.0	1406	1309.0	0.974	8.612030671	6.058897015	3.128089222
48	48) E11	0.082	0.984	97.0	1406	1309.0	0.901	6.995037483	6.058897015	3.128089222
49	48) E12	0.055	0.660	97.0	1406	1309.0	0.222	2.56962289	6.058897015	3.128089222

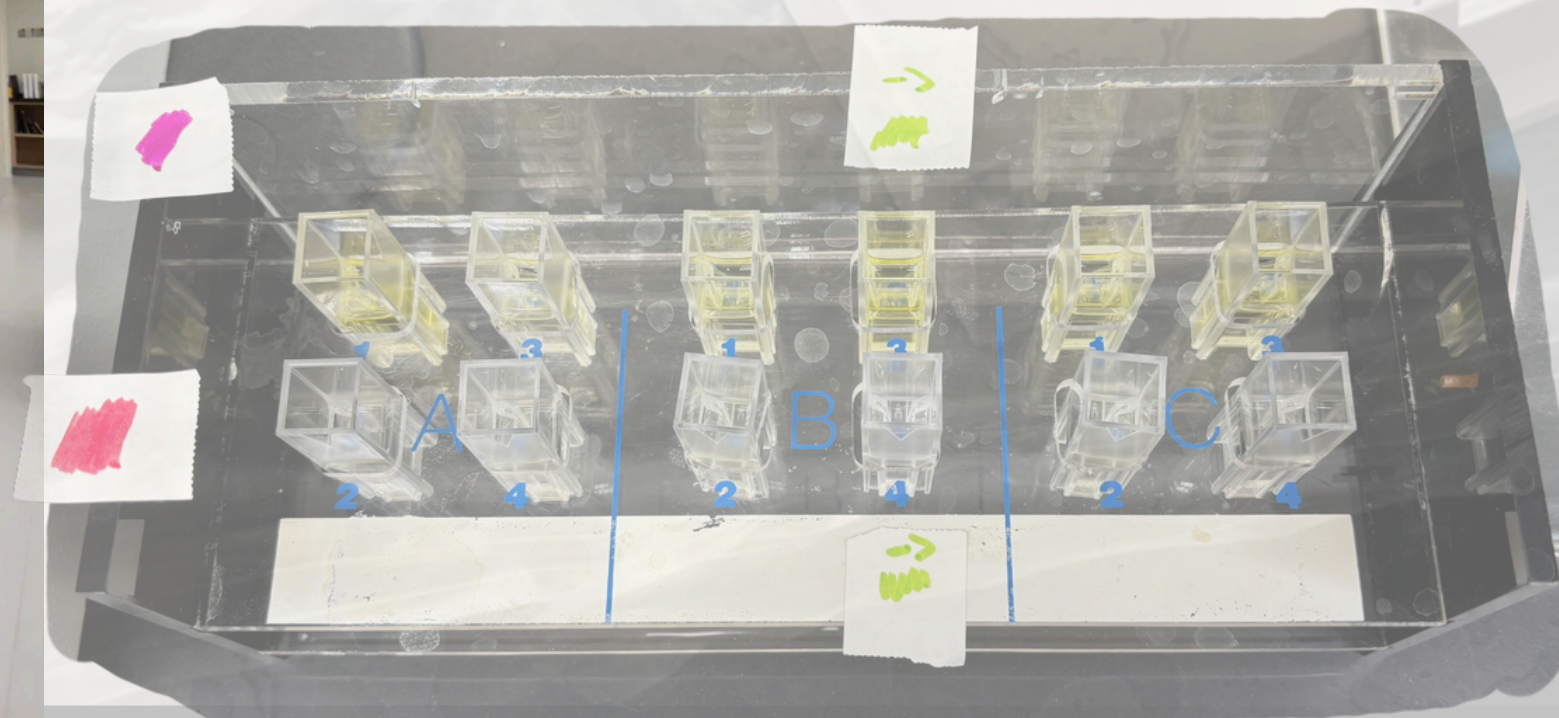
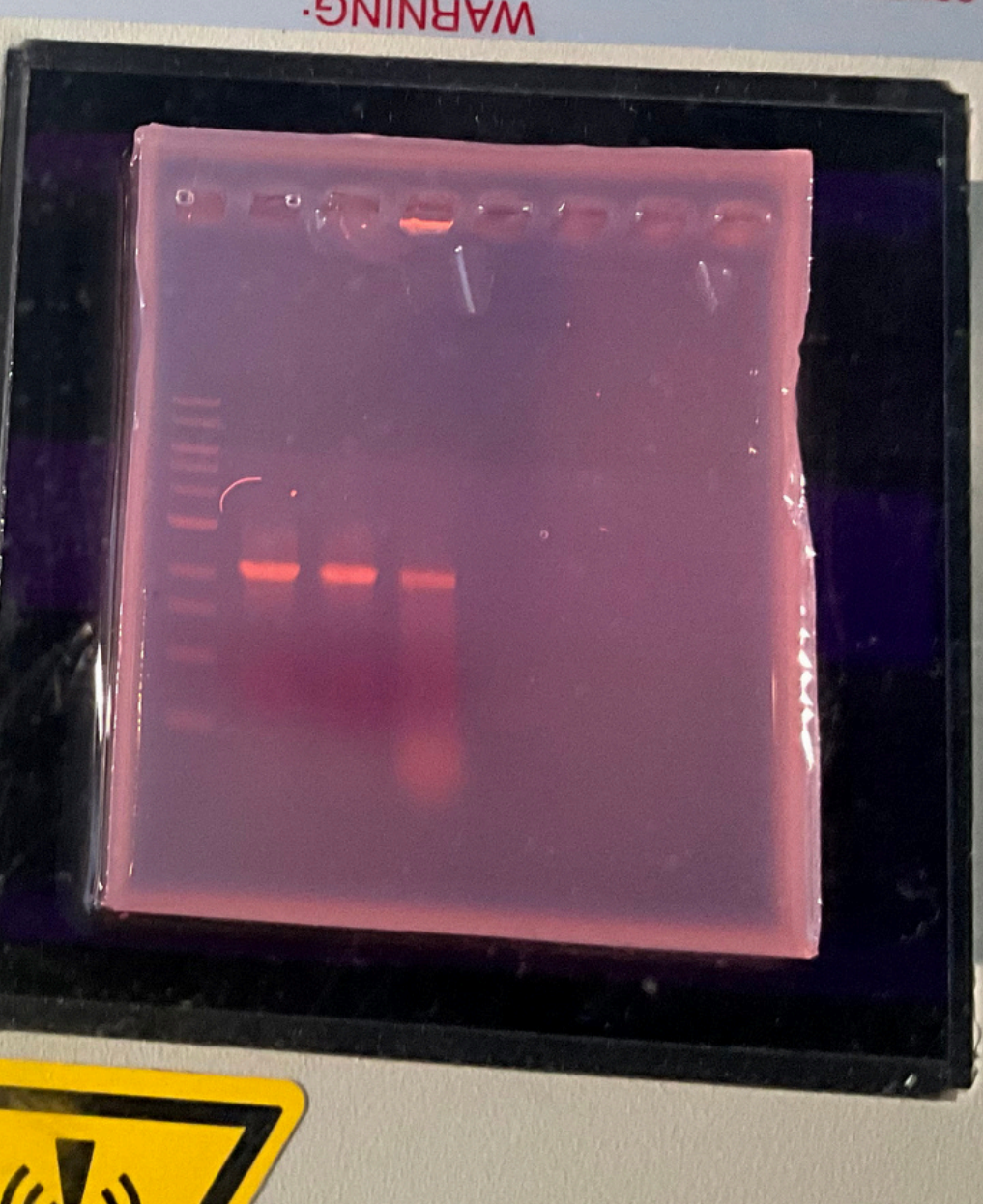
Sheet1Sheet2+



LIMITATIONS

LIMITATIONS THAT WERE PRESENT

- LIMITED TIME I HAD WITH THIS SUMMER PROJECT**
- MY KNOWLEDGE OF TERMINOLOGY WAS LIMITED WHEN STARTING A MICROBIOLOGY PROJECT.**
- BEING TRANSPARENT ON % ERROR**





WHATS NEXT ?

FALL 2024

BIOFILM ANALYSIS

LAC ASSAYS ON 4 MORE N-SOURCES

PUBLISH AND CREATE A POSTER

ASM CONFERENCE



ACKNOWLEDGMENTS



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